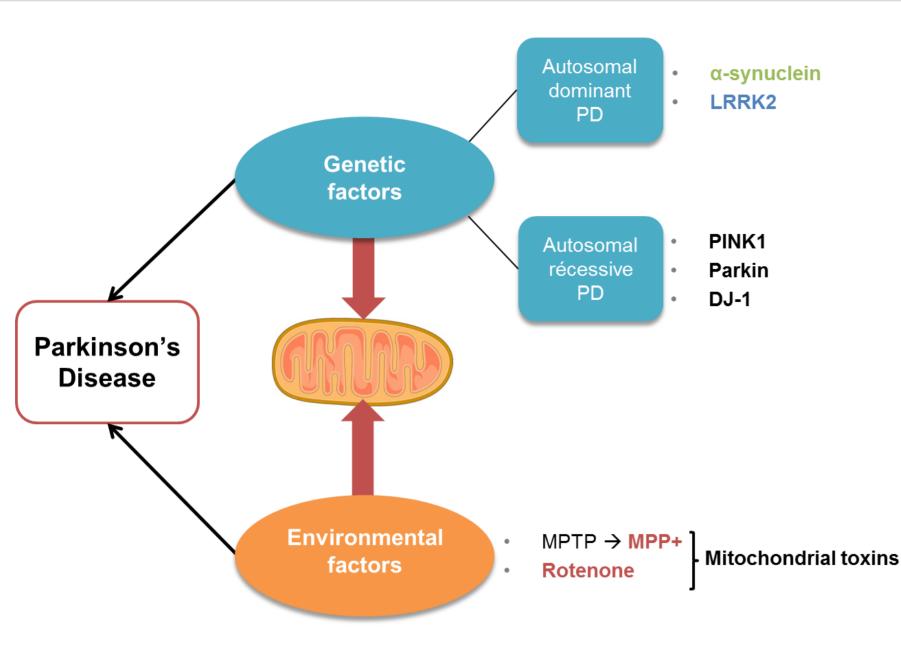
Alpha-synuclein and LRRK2's cooperation in mitochondrial dysfunctions in Parkinson's Disease (PD) BICSIGNE FLi



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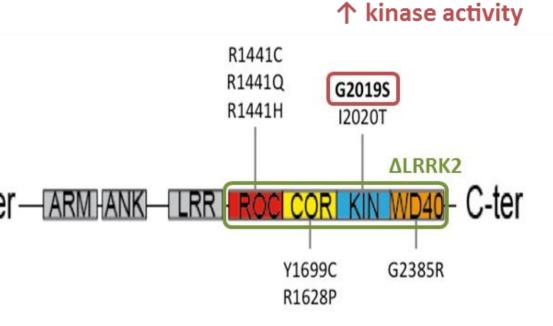
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> PD is the second most frequent neurodegenerative disease after Alzheimer's Disease. It is characterized by the selective loss of dopaminergic (DA) neurons in the SNpc and the formation of Lewy Bodies, composed by aggregates of α -synuclein (α syn). The mutation A53T of this protein can accelerate its aggregation. The pathogenesis of PD is not really understood but it results from a complex interaction between genetic and environmental factors, and many clues point towards a huge involvement of mitochondrial alterations.

LRRK2 (Leucine-Rich Repeat Kinase 2) is a protein that possesses both a kinase and a GTPase domain. It is involved in many cellular processes, such as autophagy and microtubules and neurite growth. The G2019S mutation in its kinase domain – responsible for an increased kinase activity – is the most common cause of autosomal dominant forms of PD, but is also found in N-ter ARMANK - LR ROO COR KIN WD40 some sporadic cases (Martin *et al*, 2014). We work with a truncated form of LRRK2 = Δ LRRK2, composed of the Cter part of the protein, containing the GTPase and kinase domains.



Hypothesis

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Preliminary results (Noémie Cresto, PhD)

15 weeks after the stereotaxic co-injection of an AVV coding for ΔLRRK2^{G2019S} and an AAV coding for αsyn^{A53T} in the SNpc of rats, a significant decrease of the number of DA neurons was observed compared to the injection of AAV-

PBS injected group

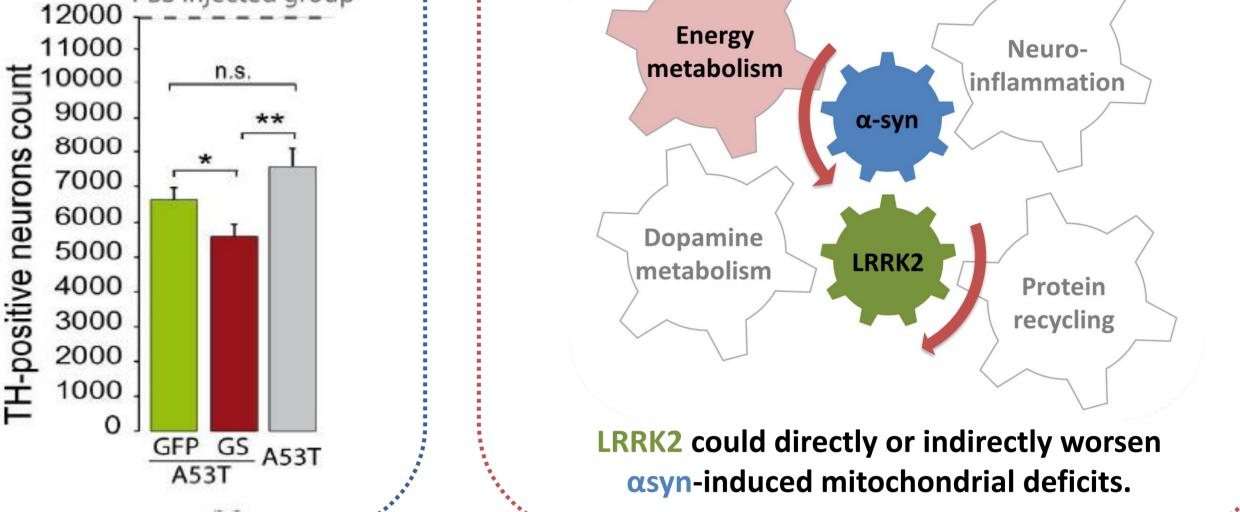
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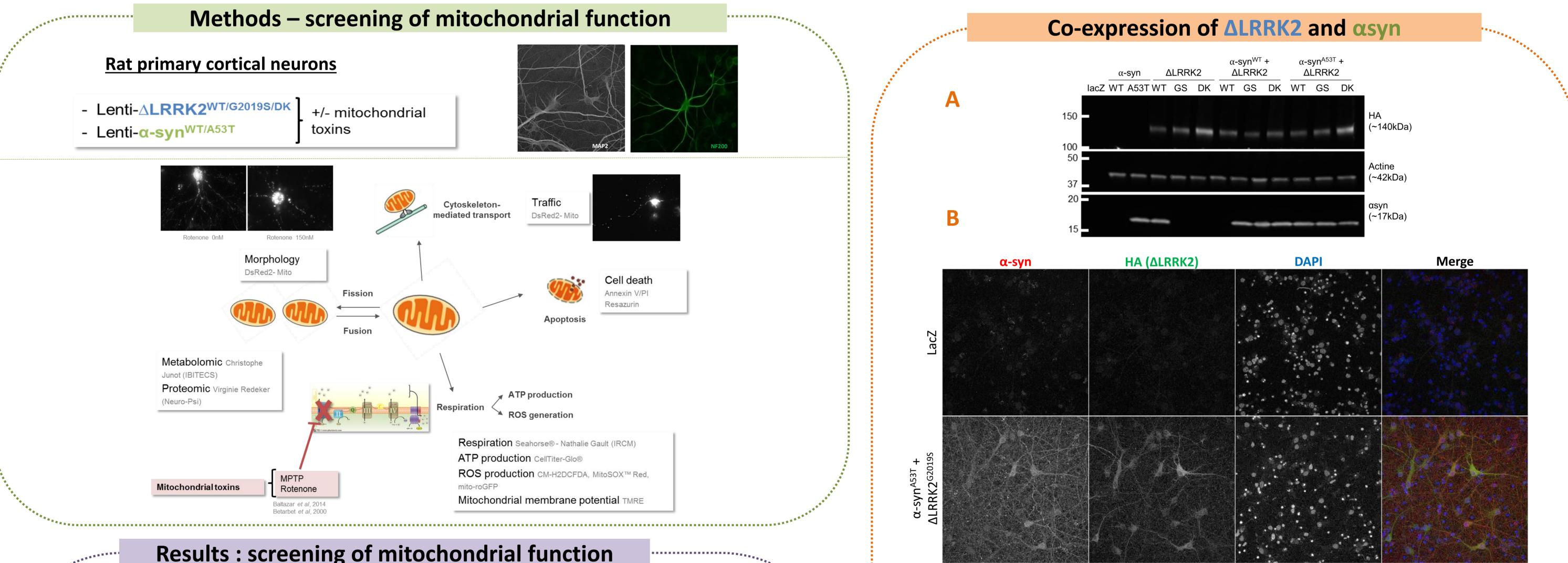
 α syn^{A53T} alone, suggesting a functional interaction between LRRK2 and α syn.

Our aim is therefore to try to decipher the mechanism of this functional interaction, focusing on the mitochondrial mechanism.

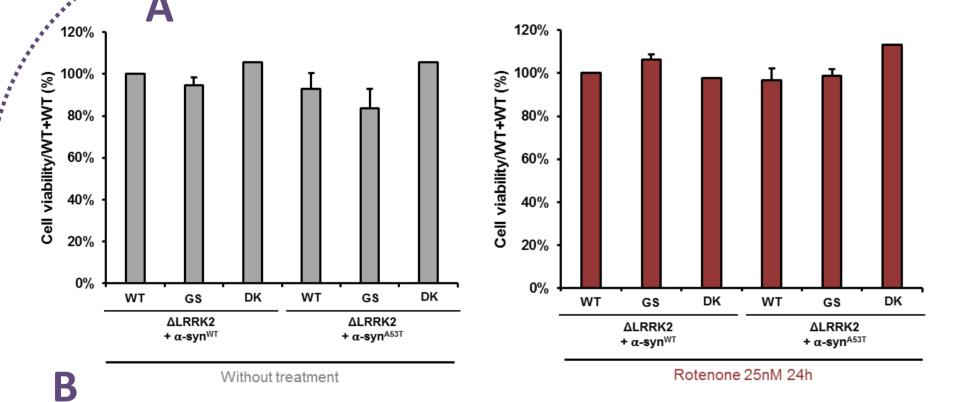
Separately, both LRRK2 and αsyn have been shown to localize to mitochondria and to alter mitochondrial function. Mitochondria is extensively studied in PD, initially because of the post-mortem description of a complex I deficiency in the striatum (Mizuno et al. 1989) and in the substantia nigra (Schapira et al. 1990) of PD patients. It is also known that environmental factors inhibiting this mitochondrial complex I (for example rotenone or MPTP) can induce parkinsonian symptoms (Langston and Ballard 1983; Schapira 2008).

 \rightarrow Mitochondrial hypothesis for the functional interaction of LRRK2 and α syn





Results : screening of mitochondrial function



120% - 100% 60% WТ WТ DK GS DK GS wт GS DK wт GS ΔLRRK2 ΔLRRK2 ΔLRRK2 ΔLRRK2 + α-syn^{WT} + α-syn^{A53} + α-syn

Primary rat cortical cultures were obtained by dissecting the cortices of E17 rat embryos. At DIV1, the cells were infected with lentivirus coding for ΔLRRK2^{WT/G2019S/DK} (20ng/100.000cells) and α syn^{WT/A53T} (10ng/100.000cells). At DIV14, cells where treated or not with 25nM of rotenone for 6h. N=1-4, n=5.

A. Cellular viability was evaluated by a resazurine test. The level of fluorescence was measured with a spectrofluorometer. The results are presented as the proportion of live cells compared to the α -syn^{WT} + Δ LRRK2^{WT} condition (mean + SEM).

B. ATP production was evaluated using the CellTiter-Glo 2.0 Assay (Promega). The level of luminescence was measured with a luminometer. The results are presented as the percentage of ATP produced compared to the α -syn^{WT} + Δ LRRK2^{WT} condition (mean + SEM). *p<0,05, ANOVA and Scheffe post hoc compared to WT+WT.

Primary rat cortical cultures were obtained by dissecting the cortices of E17 rat embryos. At DIV1, the cells were infected with lentivirus coding for lacZ or ΔLRRK2^{WT/G2019S/DK} (20ng/100.000cells) and αsyn^{WT/A53T} (10ng/100.000cells). At DIV14, the cells where either A. lysed in a triton buffer to perform a Western-Blot analysis of the expression levels of α syn and Δ LRRK2, or **B**. fixed in formaldehyde 3,7% + 5% sucrose and a double-stained for α syn and HA (for Δ LRRK2) (magnification 40X)

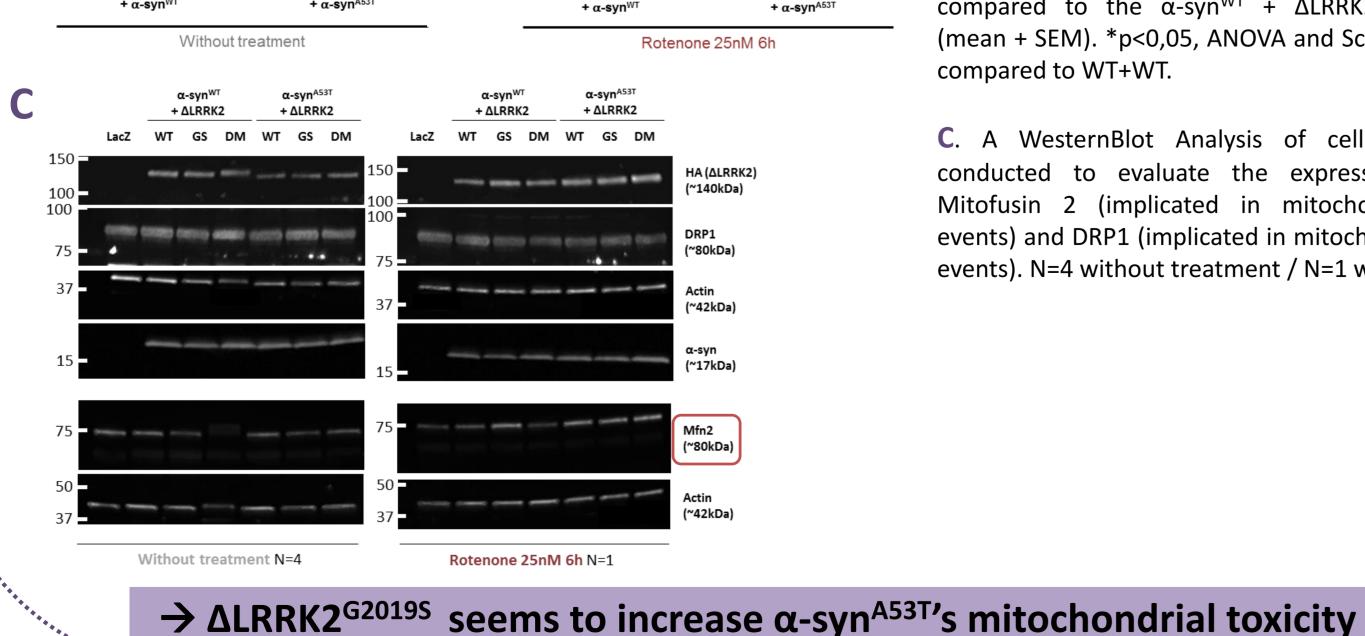
→ Validation of the cellular model of co-expression of LRRK2 and αsyn

Perspectives

Setting up a new *in vitro* model to study the mitochondrial mechanisms of the interaction between Δ LRRK2 and α syn.

Setting up the treatments to sensitize neurons to $\Delta LRRK2$ and αsyn :

- Acute drug treatment
- Chronic drug treatment



C. A WesternBlot Analysis of cell lysates was conducted to evaluate the expression level of Mitofusin 2 (implicated in mitochondrial fusion events) and DRP1 (implicated in mitochondrial fission events). N=4 without treatment / N=1 with rotenone.

Medium without antioxidants

- Screening mitochondrial function: Cell death
- Energy metabolism
- Mitochondrial traffic/morphology
- Identifying new targets: Proteomic studies Metabolomic studies

Our first results indicate that $\Delta LRRK2^{G2019S}$ modulates α -syn^{A53T}'s mitochondrial toxicity in our *in vitro* model

References

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